

‘The Gc Ms Analysis Of Ethyl Acetate Extract Of One Herbal Plant, ‘Jatrophacurcus’

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ABSTRACT

The present study deals with the GC MS analysis of ethyl acetate extracts of the aerial parts of one herbal plant ‘Jatrophacurcu’. The plant Jatrophacurcus was collected from the nearby fields at Chengalpattu, Tamil Nadu. The plant was identified by a qualified botanist at Chennai. The ethyl acetate extract of the shade dried whole plant was collected after 48 h of soaking. The extract was evaporated and the dried powder was used for GC-MS analysis by standard procedures. The GC MS profile revealed the presence of some important metabolites such as n-Hexadecanoic acid, 2-((Octan-2-yl)oxy)carbonyl)benzoic acid, Sulfurous acid, butyl heptadecyl ester, Farnesol (E), methyl ether, Palmitoleic acid, 1-Heptatriacotanol, n-Propyl 11-octadecenoate, .gamma.-Tocopherol, Fumaric acid, tetradec-3-enyl tridecyl ester. These biomolecules have many important medicinal roles, which augur well with the medicinal role of this plant.

Keywords : GC MS, Jatrophacurcus, Ethyl acetate, n-Hexadecanoic acid, 1-Heptatriacotanol, .gamma.-Tocopherol, Fumaric acid, tetradec-3-enyl tridecyl ester.

INTRODUCTION

Jatrophacurcus is a common plant available on the road side and uncultivated land, This plant was exploited for its oil as bio diesel. This plant has been traditionally used as medicine for treatment of various ailments. Crude extracts and isolated compounds from Jatrophacurcas show a wide range of pharmacological activities, such as anti-inflammatory, antioxidant, antimicrobial, antiviral, anticancer, antidiabetic, anticoagulant, hepatoprotective, analgesic and abortifacient effects. J.

curcas has been a widely used source of medicine for decades in many cultures. The present review reveals that *J. curcas* is a valuable source of medicinally important molecules and provides convincing support for its future use in modern medicine (Abdelgadir and Staden, 2013; Prasad et al, 2012; Diwan et al, 2013; Insan et al, 2013). The antibacterial activity of leaf extracts of *Jatropha curcas* was reported by Rahman et al, 2014. Othman et al, 2015 have reported the anti-inflammatory role of bioactive compounds isolated from *Jatropha curcas*. The present work reports the GC MS pattern of the ethyl acetate extracts of *Jatropha curcas*, whole plant. This is in continuation of our endeavour to establish the medicinal efficacy of the herbal and traditional systems of Ayurveda, Siddha and Unani of medicine (Priyadarshini et al, 2017; Jayakumari et al, 2017; Rao et al, 2018; Vijayalakshmi and Rao, 2019; Yuvaraj et al, 2019; Muttevi et al, 2019, Rao et al, 2019; Muttevi et al, 2020; Vijayalakshmi and Rao, 2020; Janaki et al, 2021).

MATERIALS AND METHODS

The plant *Jatropha curcas* was collected from the nearby fields at Chengalpattu, Tamil Nadu. The plant was identified by a qualified botanist at Chennai. The ethyl acetate extract of the shade dried whole plant was collected after 48 h of soaking. The extract was evaporated and the dried powder was used for GC-MS analysis by standard procedures.

GC-MS Procedure

Instrument: GC (Agilent: GC: (G3440A) 7890A. MS/MS: 7000 Triple Quad GCMS) was equipped with MS detector.

Sample Preparation

About 100 ml sample was dissolved in 1 ml of suitable solvents. The solution was stirred vigorously using vortex stirrer for 10 s. The clear extract was determined using GC for analysis.

GC-MS Protocol

Column DB5 MS (30 mm × 0.25 mm ID × 0.25 µm, composed of 5% phenyl 95% methylpolysiloxane), electron impact mode at 70 eV; helium (99.999%) was used as carrier gas at a constant flow of 1 ml/min injector temperature 280°C; auxiliary temperature: 290°C ion-source temperature 280°C.

The oven temperature was programmed from 50°C (isothermal for 1.0 min), with an increase of 40°C/min, to 170°C (isothermal for 4.0 min), then 10°C/min to 310°C (isothermal for 10 min) fragments from 45 to 450 Da. Total GC running time is 32.02 min. The compounds are identified by GC-MS Library (NIST and WILEY).

RESULTS AND DISCUSSION

The results of the GC-MS analysis of the whole plant ethyl acetate extract, along with the possible medicinal role of each molecule of *Jatropha curcas* extract are tabulated in Table 1. Figure 1 represents the GC-MS profile of ethyl acetate extract of the whole plant of *Jatropha curcas*. The identification of metabolites was accomplished by comparison of retention time and fragmentation pattern with mass spectra in the NIST spectral library stored in the computer software (version 1.10 beta, Shimadzu) of the GC-MS along with the possible pharmaceutical roles of each bio molecule as per Dr. Duke's Phytochemical and ethnobotanical data base (National Agriculture Library, USA) and others as shown in Table 1.²¹ The results as shown in Table 1 indicate the medicinal roles of some of the molecules such as n-Hexadecanoic acid, 2-((Octan-2-yloxy)carbonyl)benzoic acid, Sulfurous acid, butyl heptadecyl ester, Farnesol (E), methyl ether, Palmitoleic acid, 1-Heptatriacotanol, n-Propyl 11-octadecenoate, .gamma.-Tocopherol, Fumaric acid, tetradec-3-enyl tridecyl ester. These molecules do have medicinal roles which are supportive of the medicinal role of *Jatropha curcas*. Further work is on to establish the molecular mechanism of the medicinal roles of each molecule.

CONCLUSION

From the results it could be concluded that apart from being a source of bio-diesel manufacturing, *Jatropha curcas* has some important medicinal roles.

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REFERENCES

1. Abdelgadir, H. A., Staden, J. V. (2013) Ethnobotany, ethnopharmacology and toxicity of *Jatropha curcas* L. (Euphorbiaceae): A review. South African J of Botany, 88, 204-218
2. Prasad, D. M.R., Izam, A., Khan, M. K. R. (2012) *Jatropha curcas*: Plant of medical benefits. Journal of Medicinal Plants Research, 6(14), 2691-2699
3. Diwan, P. D., Gadhikar, Y. A., Jain, S. B. (2013) Traditional Ethnomedicinal Plants Used for Oral Health Care by Tribals of Melghat Region, Dist. Amravati (M.S.), India. Int. J. Pharm. Sci. Rev. Res., 21(1), 301-304
4. Insanu, M., Dimaki, C., Wilkins, R., et al. (2013) Rational use of *Jatropha curcas* L. in food and medicine: from toxicity problems to safe applications. Phytochemistry Reviews, 12, 107-119

5. Rahman, M. M., Ahmad, S. H., Mohamed, T. M., Rahman, Z. A. (2014) Antimicrobial compounds from leaf extracts of *Jatropha curcas*, *Psidium guajava* and *Andrographis paniculata*. *Sci World J*, 2014; Article Id 635240, 8 pages.
6. Othman, A. R., Abdullah, N., Ahmad, S. et al. (2015) Elucidation of in-vitro anti-inflammatory bioactive compounds isolated from *Jatropha curcas* L. plant root. *BMC Complement Altern Med*, 15, 11. <https://doi.org/10.1186/s12906-015-0528-4>
7. GomathiPriyadarshini, Arul Amutha Elizabeth, Jacintha Anthony, Mudiganti Ram Krishna Rao, Prabhu. K., Aiswarya Ramesh, Vani Krishna. (2017) The GC MS analysis of one medicinal plant, *Premnatomentosa*. *Journal of Pharmaceutical Sciences and Research*, **9(9)**, 1595-1597
8. Jayakumari, S., Prabhu, K., Mudiganti Ram Krishna Rao, Bhupesh, G., Kumaran, D., Aishwariya Ramesh. (2017) The GC MS Analysis of a Rare Medicinal Plant *Aloe barbadensis*. *J. Pharm. Sci. & Res.* **9(7)**, 1035-1037
9. Rao, M. R. K., Vijayalakshmi, N. (2018) Preliminary phytochemical and GC MS analysis of different extracts of *Sphaeranthus indicus* leaves. *Indo American J of Pharmaceuical Sciences*, **5(3)**, 1511-1520
10. Vijayalakshmi, N., Mudiganti Ram Krishna Rao. (2019) The antioxidant studies of two medicinal plants, *Sphaeranthus indicus* and *Psophocarpus tetragonolobus*. *Asian J of pharmaceutical and Clinical Res*, **12(1)**, 321-327.
11. Yuvaraj, R., Mudiganti Ram Krishna Rao, Prabhu, K., Lakshmisundram, R., SampadShil, Sathish Kumar, M., Vijayalakshmi, N. (2019) The GC MS study of one medicinal plant, *Stachytarpheta indica*. *Drug Invention Today*, **12(9)**, 1665-1669
12. Muttevi Hyageva Kumar, Prabhu, K., Mudiganti Ram Krishna Rao, Lakshmisundram, R., SampadShil, Sathish Kumar, M., Vijayalakshmi, N. (2019) The GC MS study of one medicinal plant, *Dodonaea angustifolia*. *Drug Invention Today*, **12(9)**, 1661-1664
13. Mudiganti Ram Krishna Rao, Vijayalakshmi, N., Prabhu, K., Sathish Kumar, M. (2019) The gas chromatography–mass spectrometry study of *Moringa oleifera* seeds. *DIT*, **12(10)**, 2172-2175
14. Muttevi Hyageva Kumar, Prabhu, K., Mudiganti Ram Krishna Rao, Lakshmisundram, R., SampadShil, Sathish Kumar, M., Vijayalakshmi, N. (2020) The GC MS study of one medicinal plant, *Aristolochia indica*. *DIT*, **12(12)**, 2919-2923.

15. Vijayalakshmi, N., Mudiganti Ram Krishna Rao. (2020) 'Preliminary phytochemical and antioxidant studies of leaf extracts of one medicinal plant, Vitexnegundo".RJPT, 13(5), 2167-2173

16. Janaki C. S., Prabhu K., Mudiganti Ram Krishna Rao,VenkatRamaiah, ShrutiDinkar, Vijayalakshmi, N., Kalaivannan. J. (2021) The GC MS analysis of Ethyl acetate extract of Merremiaemerginata'.Ind J of Natural Sciences, 12(67), 33638-33646

17. Dr.Duke's Phytochemical and Ehnobotanical Databases.U.S. Department of Agriculture, Agricultural Research Service.1992-2016. Dr. Duke's Phytochemical and Ethnobotanical Databases. Home Page, <http://phytochem.nal.usda.gov/> <http://dx.doi.org/10.15482/USDA.ADC/1239279>

Figure 1. Shows the GC MS profile graph of ethyl acetate extract ofJatrophacurcus

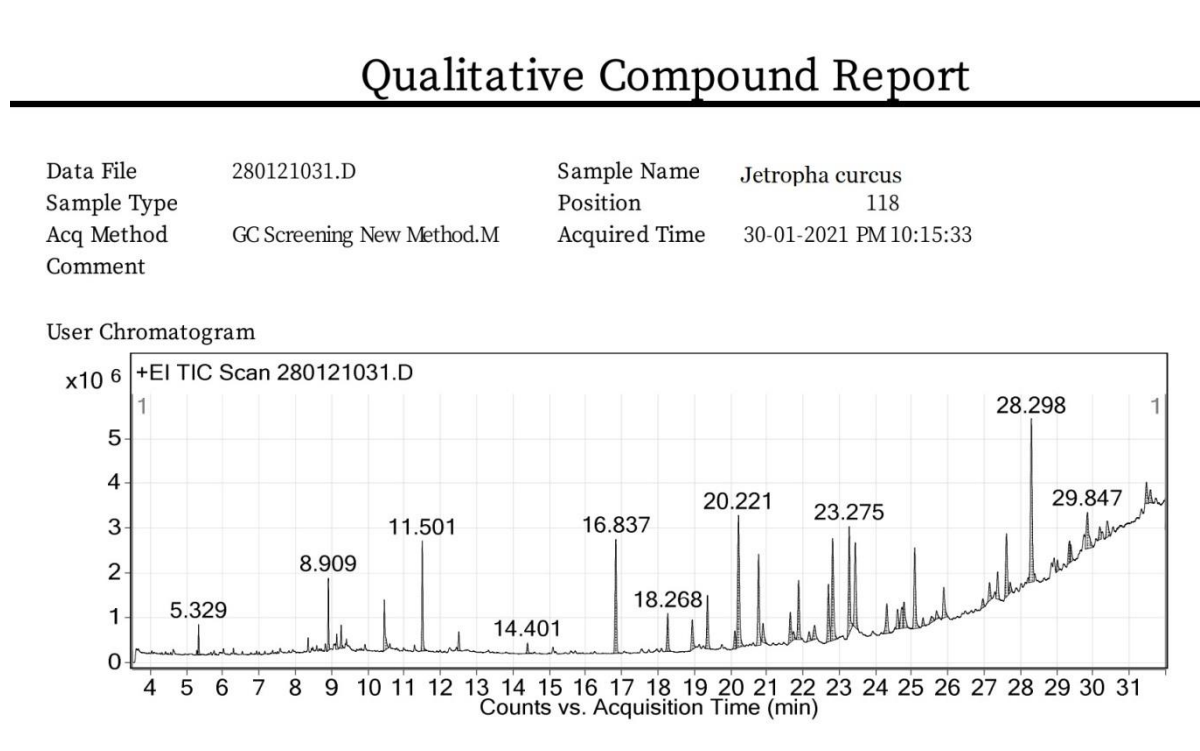


Table1. Indicates the retentions time, types of possible compound, molecular formula, molecular mass, percentage peak area and the possible medicinal roles of each compound as shown in the GC MS profile of Jatrophacurcus

Ret. Time	Compound	Mol. formula	Mol. Mass	% Peak Area	Possible Medicinal Role
8.91	Bicyclo[3.1.1]heptane, 2,6,6-trimethyl-	C ₁₀ H ₁₈	138.1	2.04	Not Known
10.45	n-Hexadecanoic acid	C ₁₆ H ₃₂ O ₂	256.2	2.50	Acidifier, Arachidonic acid Inhibitor, Increases Aromatic Amino acid decarboxylase activity, Inhibits production of uric acid, Urine acidifier, Anaphylactic, Arylamine N acetyltransferase inhibitor, decreases norepinephrine production, Down regulates nuclear and cytosol androgen reuptake, GABA-nergic, Increase NK cell activity, inhibits production of tumor necrosis factor, Myo-neuro-stimulator
11.50	Cyclohexanol, 5-methyl-2-(1-methylethyl)-, (1.alpha.,2.beta.,5.alpha.)-(./-.)-	C ₁₀ H ₂₀ O	156.2	4.27	Not Known
16.84	Dodecane, 1-fluoro-	C ₁₂ H ₂₅ F	188.2	6.24	Not Known
18.27	2-((Octan-2-yl)oxy)carbonyl)benzoic acid	C ₁₆ H ₂₂ O ₄	278.2	2.20	Acidifier, Arachidonic acid inhibitor, Increases Aromatic Amino acid Decarboxylase activity
19.37	Sulfurous acid, butyl heptadecyl ester	C ₂₁ H ₄₄ O ₃ S	376.3	3.25	Acidifier, Arachidonic acid inhibitor, Increases Aromatic Amino acid Decarboxylase activity
20.22	Farnesol (E), methyl ether	C ₁₆ H ₂₈ O	236.	9.68	anticancer, antidote, antitumor,

			2		Cytochrome-P450-2E1-Inhibitor, Decreases C-Teleopeptide Excretion, Decreases Deoxyypyridinoline Excretion, Decreases Endothelial Leukocyte Adhesion, Decreases Epinephrine Production, Decreases Oxalate Excretion
20.77	Palmitoleic acid	C16H30O2	254. 2	5.57	Acidifier, Arachidonic acid inhibitor, Increases Aromatic Amino acid Decarboxylase activity
22.32	1-Heptatriacotanol	C37H76O	536. 6	1.21	Antibacterial, anticancer, antiprotozoal, chemo-preventive, anti-inflammatory, antimalarial, anti-flu, antiviral, enzyme inhibitor, anti-hyper-cholesterolemic
22.70	Butyl 9-hexadecenoate	C20H38O2	310. 3	3.73	Not Known
22.82	n-Propyl 11-octadecenoate	C21H40O2	324. 3	7.45	Anaphylactic, Antitumor, Arylamine-N-Acetyltransferase- Inhibitor, Decreases Norepinephrine Production, Down regulates nuclear and cytosol androgen reuptake, GABA-nergic, Increases natural killer cell activity, Inhibits Production of Tumor Necrosis Factor, Myo-neuro- stimulant, N-Cholinolytic, NADH- Oxidase-Inhibitor, NADH- Ubiquinone-Oxidoreductase- Inhibitor
23.44	1-Nonylcycloheptane	C16H32	224. 3	5.46	Not known

24.61	Octacosyl acetate	C30H60O2	452. 5	1.35	Not known
24.74	.gamma.-Tocopherol	C28H48O2	416. 4	1.79	Tocopherol synergist, PPAR-gamma antagonist
25.08	9-Octadecenoic acid (Z)-, 2-(acetyloxy)-1-[(acetyloxy)methyl]ethyl ester	C25H44O6	440. 3	5.60	Not known
25.88	1,19-Eicosadiene	C20H38	278. 3	2.29	Not known
27.15	Fumaric acid, tetradec-3-enyl tridecyl ester	C31H56O4	492. 4	1.27	Acidifier, Arachidonic acid inhibitor, Increases Aromatic Amino acid Decarboxylase activity