

The Molecular Variation Of Wzx and Wzy Genes In Multi Drugs Resistant *Pseudomonas aeruginosa* Isolated From Wounds

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Abstract

Pseudomonas aeruginosa infection is one of the health problems in hospital patients, the multi-drug resistance isolates were enrolled in the present study to detect genetic variation of some B-band LPS synthesis genes, the *wzx* and *wzy* genes were detected using monoplex-PCR, the results show that the *wzx* was detected in 87.5% of isolates while it wasn't found in 12.2%, the *wzy* gene was found in 50% of isolates, the other variations were found both genes (*wzx+wzy*) in 50%, and 37.5% of isolates have *wzx* without *wzy*, and 12.5% have *wzy* without *wzx*. The present study concluded that the genetic variants of B-band (*wzx* and *wzy*) involvement in the wound infections and multidrug resistance of *P. aeruginosa*.

Key words: Molecular Variation, *wzx* and *wzy* Genes, Multi Drugs Resistance, *Pseudomonas aeruginosa*, Wound

Introduction

The increments of bacterial infections in Iraq in the last years and the high resistance to a wide range of antibiotics, in addition to environment pollutions, and lack of health awareness, all these factors led to increase infection rates of different bacterial isolates (Al-Kadmy et al., 2019). The pathogenic bacteria characterized by an elaborate assortment of cell-associated and extracellular bacterial substances for utilizing in the establishment of infection and colonization with a host (Peterson et al., 1996). One of these productions is Lipopolysaccharide (LPS) molecules, its protective factor against the lysis activity of hosts, the most LPS heterogeneous part is O antigen, moreover, this region is conferred serum resistance to the organism (Farhana and Khan, 2021). The genetic variation of O antigen based on the genetic polymorphisms of the *rfp* genes cluster encoded to enzymes utilized in the O-antigen synthesis and construction (Tarr et al., 2000), these genes encoded to enzymes involved in the sugar synthesis, construction of sugar and O subunit transferase enzyme, and proteins utilized in the construction of O antigen by subunits assembly, these genes are *wzx* and *wzy* which encoded to transporter of O-antigen or flippase and O-antigen polymerase respectively that proved by some studies in different bacterial species (Samuel and Reeves, 2003; Feng et al., 2004; Vinés et al., 2005; Patel et al., 2012).

In some studies two theories were suggested to explain the lowering C+G percent of heteropolymeric O-antigen encoded gene clusters. The first theory proposed the lateral transfer of genetic materials from lower C+G percent organism (Reeves, 1993, Schnaitman and Klena, 1993). The second theory, dependent on the phylogenetic analysis, utilized the mechanisms of potential translational regulation, regarding to the atypical molar ratio is impacted in the codons utilized atypical for the background. The intriguing schism between the C+G percent of interdependent homopolymeric and heteropolymeric O-antigen clusters in the same backgrounds lends credence to the second theory (Rocchetta et al., 1999; Geue et al., 2017).

According to the O antigen role in the *Pseudomonas aeruginosa* infections and serotyping, the present study suggested a molecular assessment of *wzx* and *wzy* in several isolates collected from wound infection in hospitalized patients.

Methodology

Pseudomonas aeruginosa isolates that isolated from wound infection of hospitalized patients were enrolled in the present study, these isolates were diagnosed by macroscopic and microscopic tests, and antibiotics sensitivity was applied, then DNA was isolated from each isolate and electrophoresis were performed for DNA profile of bacteria (Mona et al., 2019). The *wzx* and *wzy* detected by monoplex-PCR using the primers *wzx* F CCG, GGT, TTC, GAT, TTG, TGA, AGG, TTG. R CAC, AAC, AGC, CAC, TAC, TAG, GCA GAA, *wzy* F GAA, ATT, ATG, CCA, TCT, TGG, CGA, GCG. R CAT, GTG, AAG, CCT, GAA, GGC, AAA, CTC. Using thermo cycler (biometra, German) 94°C for 5 min, 31 cycles including (94°C 30 s, 58°C 30 s, 72°C 30 s) finally 72°C for 10 min, The amplification sizes *wzx* 255 bp and *wzy* 450 bp which visualized using 1% agarose, 0.5 TBE, 100 V for 30 min (Raad et al., 2021; Al-Terehi et al., 2018).

Results and Discussion

The present output shows that all isolates didn't contain plasmids, the Genetic Variation of *wzx* and *wzy* genes show that the *wzx* was detected in 87.5% of isolates while didn't found in 12.2%, the *wzy* gene found in the 50% of isolates, the other variations were found both genes (*wzx*+*wzy*) in 50%, and 37.5% of isolates have *WZX* without *wzy*, and 12.5% have *wzy* without *wzx* (figure 1). The amplification products for both genes are shown in figure (2).

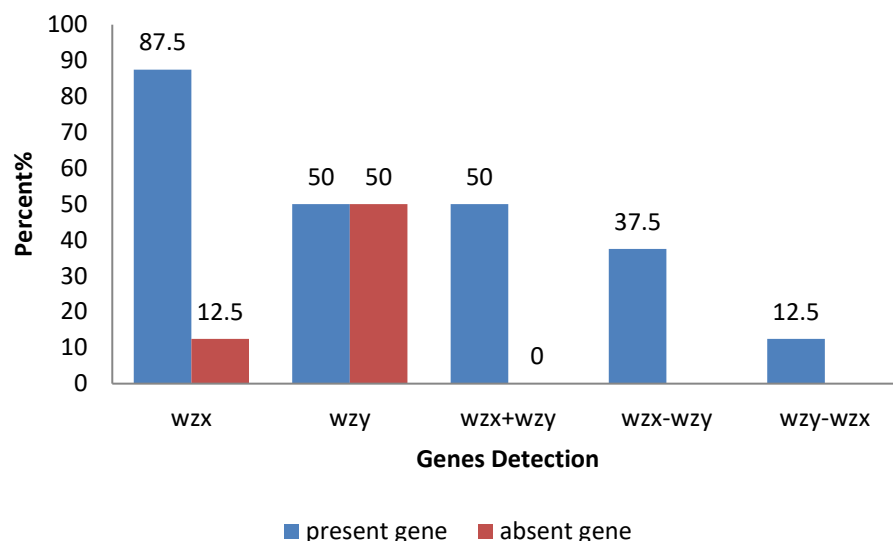


Figure (1) the detection percent's of wzx and wzy Genes in pseudomonas aeruginosa isolates

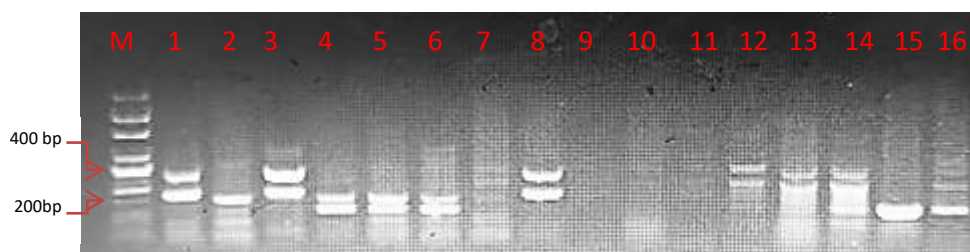


Figure (2) the amplification products of wzx and wzy Genes in pseudomonas aeruginosa isolates, lane M DNA marker (11-1000bp), lanes 1-8 wzx amplification products (255 bp), lanes 9-16 wzy amplification products (450 bp).

The bacterial infection of the wound may lead to severe complications especially with the multidrug resistance isolates of *Pseudomonas aeruginosa*, thus the detection of the virulence factor structure should be studied for drug design and therapy (Al-Terehi et al., 2021).

The O antigen has a major role in the serological distinction, which is one of the most components of LPS that present in different *G^{-ve}* bacteria, moreover LPS is thermostable and variant among species (Maldonado et al., 2016). The variant of LPS based on some genetic polymorphisms that encoded to the enzymes and proteins contributed in the LPS synthesis, in the present study wzx and wzy Genes show different percent of present these genes in *Pseudomonas aeruginosa* isolated from wounds, these differences may be responsible for the virulence of isolates which might be impacted on the LPS structure that have a role in stimulating the robust pro-inflammatory of the immune system in mammals (Lam et al., 1992).

In *Pseudomonas aeruginosa* the Genes involved in LPS biosynthesis classified into three groups A-band LPS, B-band LPS synthesis, Core oligosaccharide synthesis and housekeeping function impacting in the synthesis of LPS wzx and wzy under B-band LPS synthesis (Rocchetta et al., 1999), another study found 61.53% of *P. aeruginosa* isolates production rough LPS and

serotype O11 LPS when transformed with O-antigen gene (rfb). Therefore the wbp region has acquired mutations, these mutations responsible on the changes in LPS phenotype (Evans et al. 1994).

Other results show two bands of wzx and wzy amplification products, these amplifications may be non-specific amplifications, or two copies of the gene, although of the positive and negative control were applied. Also, it may be resulted from the genetic variations of genes among isolates that produced ranging in amplification size products (Naji Hasan, R. & Abdal Kareem Jasim, S. (2021); Goldberg et al., 1992). On the other hand, the present isolates were multi-resistance drugs, and this resistance belongs to the genetic variants in the B-band encoded genes, the antibiotic treatment effective of *P. aeruginosa* infections still health problems, this belongs to the high intrinsic resistance to antimicrobial substances which resulted from the outer membrane low permeability (Hancock et al., 1998). The present study concluded that the genetic variants of B-band (wzx and wzy) involvement in the wound infections and multidrug resistance of *P. aeruginosa*.

References

- 1- Farhana A, Khan YS. Biochemistry, Lipopolysaccharide. [Updated 2021 Apr 29]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2021 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK554414/>.
- 2- Tarr PI, Schoening LM, Yea YL, Ward TR, Jelacic S, et al. (2000) Acquisition of the rfb-gnd cluster in evolution of *Escherichia coli* O55 and O157. *J Bacteriol* 182: 6183–6191. PMID:11029441.
- 3- Samuel G, Reeves P (2003) Biosynthesis of O-antigens: genes and pathways involved in nucleotide sugar precursor synthesis and O-antigen assembly. *Carbohydr Res* 338: 2503–2519. PMID:14670712.
- 4- Liu, D., R. A. Cole, and P. R. Reeves. (1996). An O-antigen processing function for Wzx (RfbX): a promising candidate for O-unit flippase. *J. Bacteriol.* 178:2102-2107. [PMC free article] [PubMed] [Google Scholar].
- 5- Mona, A.-T.N., Raad, H.N., Methak, A.-J.J., (2019). In silico design bacterial expression vectors of therapeutic strategies of lethal-micro-RNA-7. *Research Journal of Biotechnology*, 2019, 14(Special Issue I), pp. 38–47
- 6- Fitzgerald, C., Sherwood, R., Gheesling, L. L., Brenner, F. W., & Fields, P. I. (2003). Molecular analysis of the rfb O antigen gene cluster of *Salmonella enterica* serogroup O:6,14 and development of a serogroup-specific PCR assay. *Applied and environmental microbiology*, 69(10), 6099–6105. <https://doi.org/10.1128/aem.69.10.6099-6105.2003>.
- 7- Raad Naji Hasan, Saade Abdal Kareem Jasim, Yasameen Hassan Ali, (2021). Detection of fimH, kpsMTII, hlyA, and traT genes in *Escherichia coli* isolated from Iraqi patients with cystitis. *Gene Reports*, Volume 26, 2022, 101468, <https://doi.org/10.1016/j.genrep.2021.101468>. (<https://www.sciencedirect.com/science/article/pii/S2452014421004520>)
- 8- Al-Terehi M, Khazaal S, Muhammed H, Behjet R. (2018). Molecular Detection of wzx1 and wzy Genes in Multi Drugs Resistance *E. coli* isolates. *International Journal of Pharmaceutical Quality Assurance* 2018; 9(3); 304-307

- 9- Al-Terehi, M.N., Al-Jboory, M.J., Hasan, R.N. (2021). DIFFERENT PATTERNS OF INSERTION REGULATORY SEQUENCES IN GENETIC ENGINEERING AND GENE THERAPY VECTORS. *Biochemical and Cellular Archives*, 2021, 21, pp. 2057–2063
- 10- Maldonado RF, Sá-Correia I, Valvano MA. Lipopolysaccharide modification in Gram-negative bacteria during chronic infection. *FEMS Microbiol Rev.* 2016 Jul;40(4):480-93. [[PMC free article](#)] [[PubMed](#)] [[Reference list](#)].
- 11- Rocchetta, H. L., Burrows, L. L., & Lam, J. S., (1999). Genetics of O-antigen biosynthesis in *Pseudomonas aeruginosa*. *Microbiology and molecular biology reviews* : MMBR, 63(3), 523–553. <https://doi.org/10.1128/MMBR.63.3.523-553.1999>
- 12- Evans D J, Pier G B, Coyne M J, Jr, Goldberg J B. The rfb locus from *Pseudomonas aeruginosa* strain PA103 promotes the expression of O antigen by both LPS-rough and LPS-smooth isolates from cystic fibrosis patients. *MolMicrobiol.* 1994;13:427–434. [[PubMed](#)] [[Google Scholar](#)].
- 13- Lam J S, Graham L L, Lightfoot J, Dasgupta T, Beveridge T J. Ultrastructural examination of the lipopolysaccharides of *Pseudomonas aeruginosa* strains and their isogenic rough mutants by freeze-substitution. *J Bacteriol.* 1992;174:7159–7167. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)].
- 14- Naji Hasan, R., Abdal Kareem Jasim, S. (2021). 'Detection of Pantone-Valentine leukocidin and MecA Genes in *Staphylococcus aureus* isolated from Iraqi Patients', *Archives of Razi Institute*, 76(4), pp. 1054-1059. doi: 10.22092/ari.2021.355962.1751
- 15- Goldberg J B, Hatano K, Meluleni G S, Pier G B. Cloning and surface expression of *Pseudomonas aeruginosa* O antigen in *Escherichia coli*. *Proc Natl Acad Sci USA.* 1992;89:10716–10720. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)].
- 16- Hancock R E W. Resistance mechanisms in *Pseudomonas aeruginosa* and other nonfermentative gram-negative bacteria. *Clin Infect Dis.* 1998;1:S93–S99. [[PubMed](#)] [[Google Scholar](#)].
- 17- Reeves P. Evolution of *Salmonella* O antigen variation by interspecific gene transfer on a large scale. *Trends Genet.* 1993;9:17–22. [[PubMed](#)] [[Google Scholar](#)].
- 18- Schnaitman C A, Klena J D. Genetics of lipopolysaccharide biosynthesis in enteric bacteria. *Microbiol Rev.* 1993;57:655–682. [[PMC free article](#)] [[PubMed](#)] [[Google Scholar](#)].
- 19- Al-Kadmy IMS, Ibrahim SA, Al-Saryi N, Aziz SN, Besinis A, Hetta HF. Prevalence of Genes Involved in Colistin Resistance in *Acinetobacter baumannii*: First Report from Iraq. *Microb Drug Resist.* 2020 Jun;26(6):616-622. doi: 10.1089/mdr.2019.0243. Epub 2019 Dec 9. PMID: 31816255.
- 20- Peterson JW. Bacterial Pathogenesis. In: Baron S, editor. *Medical Microbiology*. 4th edition. Galveston (TX): University of Texas Medical Branch at Galveston; 1996. Chapter 7. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK8526/>
- 21- Geue Lutz, Menge Christian, Eichhorn Inga, SemmlerTorsten, WielerLothar H., Pickard Derek, Berens Christian, Barth Stefanie A. Evidence for Contemporary Switching of the O-Antigen Gene Cluster between Shiga Toxin-Producing *Escherichia coli* Strains Colonizing Cattle, *Frontiers in Microbiology*, 2017, 424, DOI=10.3389/fmicb.2017.00424.
- 22- Feng, L., Senchenkova, S. N., Yang, J., Shashkov, A. S., Tao, J., Guo, H., Cheng, J., Ren, Y., Knirel, Y. A., Reeves, P. R., & Wang, L. (2004). Synthesis of the heteropolysaccharide O

antigen of Escherichia coli O52 requires an ABC transporter: structural and genetic evidence. Journal of bacteriology, 186(14), 4510–4519.
<https://doi.org/10.1128/JB.186.14.4510-4519.2004>