

In Vitro Rumen Methanogenesis Inhibition Ability Of Brown Seaweed From Nusakambangan Coast, Cilacap, Indonesia

Caribu Hadi Prayitno¹, Nur Hidayat², Munasik³

Faculty of Animal Science, Jenderal Soedirman University, Indonesia

Abstract The objective of this study was to examine the effects of Sargassum sp. brown seaweed supplementation on methane gas production, methanogen, and total gas as the main indicator of anti methanogenesis agent. The materials used in this study: Sargassum sp. brown seaweed meal (from Nusakambangan Coast, Cilacap, Indonesia), sheep rumen fluid, and laboratory equipment. Completely Randomized Design (CRD) was used in the experimental research of 6 treatments, each with four replications. The applied treatments were P0 = rumen fluid with basal feed (CP = 11.8% and TDN = 60%); P1 = P0 + 0.03% monensin; P2 = P0 + 1% Sargassum sp.; P3 = P0 + 2% Sargassum sp.; P4 = R0 + 3% Sargassum sp.; and P5 = R0 + 4% Sargassum sp. The result of the experiment showed that the supplementation of Sargassum sp. decreased total gas, methane gas concentration and methanogen, increased dry matter digestibility and organic matter digestibility, and did not significantly affect (P>0.05) protozoa. Supplementation of 2% Sargassum sp. brown seaweed meal save 82.95% and decreased methanogen by 75.17%. In conclusion, Sargassum sp. brown seaweed meal was an effective anti methanogenesis agent.

Keywords Sargassum sp., Anti-methanogenesis, Methanogen

1. Introduction

Global warming issue has been the concern of the global citizen because of the severe impact on human beings. Methane is a gas that contributes to global warming with 28-fold strength compared to CO₂. Some researchers from the livestock sector have attempted to reduce methane emission especially from the ruminants [1,2,3]. Methanogenesis in ruminant cattle is due to carbohydrate fermentation which turns into volatile fatty acid (VFA), CO₂ and H₂ [4]. CO₂ and H₂ resulted from fermentation are converted into methane by the methanogen. The present study investigated the potential of brown seaweed Sargassum sp as an anti-methanogenesis agent in sheep cattle. Phlorotannin in Sargassum sp will bind the methanogen protein-enzyme which then lyses the membrane cell of methanogen. This approach was conducted in vitro to identify the ability of Sargassum sp. in inhibiting methanogenesis through the concentrate of total gas, methane gas, methanogen population, protozoa population and the end product of fermentation.

2. Materials and Methods

The materials used in this study were the ruminal fluid of sheep that had been adapted with Sargassum sp. seaweed meal for 1.5 months, Sargassum sp. seaweed meal, 0.03% monesin, field grass, concentrate as basal feed with 60:40 forage and concentrate. The equipment used consisted of a measurement set for total gas and methane gas (0,2,4,8,12,18,24,30,36 and 48 hours) that described by Inthapanya et al [5], methanogen [6], optical microscope model CX23LEDRFS1, counting chamber Sedgewick-Rafter, incubator, and autoclave Prestige Medical 2100 Classic. Media Paynter and Hungate [7] (per 100 ml: 1.3 g agar; 0.05g KH₂PO₄; 0.05 g K₂HPO₄; 0.05 g (NH₄)₂ SO₄; 0.10 g NaCl; 0.05 g MgSO₄; 0.013 g CaCl₂.2H₂O; 0.1% 0,1 ml resazurin solution, and 70 ml aquadest, 30 ml rumen liquid, 0.5 g NaHCO₃, 0.048 g Na₂SO₃ and 0.03 g L-metionin HCl), and ingredients for protozoa coloring (methylgreen formalin saline, per 100 ml 35% formaldehyde solution 10 ml; 90 ml aquadest; 0.03 g methyl green and 0.8 g NaCl).

No	Feedstuff	PO	P1	P2	Р3	P4	P5
			%				
1	Field grass	60	60	60	60	60	60
2	Concentrate	40	40	40	40	40	40
	- Tapioca by-product	47.5	47.5	47.5	47.5	47.5	47.5
	- Pollard	25	25	25	25	25	25
	- Coconut waste	7.72	7.72	7.72	7.72	7.72	7.72
	- Palm waste	11	11	11	11	11	11
	- Soybean meal	1.57	1.57	1.57	1.57	1.57	1.57
	- Limestone	1.43	1.43	1.43	1.43	1.43	1.43
	- Salt	1	1	1	1	1	1
	- Molasses	4.5	4.5	4.5	4.5	4.5	4.5
	- Mineral	0.28	0.28	0.28	0.28	0.28	0.28
3	Sargassum sp.	0	0	1	2	3	4
4	Monensin	0	0.03	0	0	0	0

Table 1. The composition of the experimental ration

A completely randomized design was used for six treatments each with four replicates.

The treatments consisted of P0 = basal ration (CP = 11.8% and TDN = 60%), P1 = P0 + monensin 0.03\%, P2 = P0 + Sargassum sp. 1\%, P3 = P0 + Sargassum sp. 2\%, P4 = P0 + Sargassum sp. 3\%, P5 = P0 + Sargassum sp. 4\%.

The data were analyzed using variant analysis with a completely randomized design. The significantly different result were further analyzed using Duncan multiple range test (DMRT) at the levels of 5% and 1% [8].

3. Findings and Discussion

The supplementation of Sargassum sp. Seaweed to sheep feed significantly increased the concentration of total gas. The effect of 48h observation indicated that P2 treatment (2% Sargassum sp.) was the most

optimum treatment to inhibit the production of total gas. Ruminal carbohydrate fermentation produced several gasses including CO2, CH4, and H2 [3,9]. In contrast, Belanche et al [10] reported that 2% Ascophyllum nodosum (ASC) seaweed did not affect the total gas.

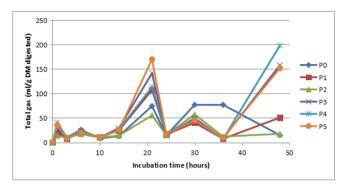


Figure 1. Total gas concentration during incubation time (0-48 h)

The result of this study showed that supplementation of 2% Sargassum sp in sheep feed observation at 4 hours decreased total gas by 25.76% and at the end observation (48 hours) decreased 11.02% of total gas. Level of Sargassum sp above 2% provide a response to total gas that varies. The supplementation of Sargassum sp. seaweed significantly affected the decrease of methane gas concentration. Furthermore, P2 treatment (2% Sargassum sp.) across the incubation hours (0,2, 4, 8,12, 18,24,30,36 and 48 hours) was the optimum level to inhibit methane gas production. The result showed that supplementation Sargassum sp. of 2% decreased methane gas by 88.83% at the 8th hour of observation and at the end of the observation period (48 hours) decreased 82.95%. The ability of Sargassum sp. to reduce methane gas was stronger than monensin (P1). The presence of phlorotanin from Sargassum sp. was thought to be able to suppress methanogen activity.

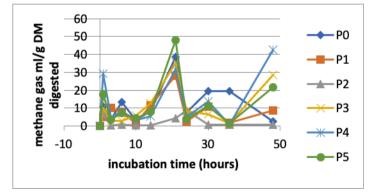


Figure 2. Methane gas concentration during incubation time (0-48h)

Mean and standard deviation of the total protozoa and methanogen and protozoa supplemented with Sargassum sp. are presented in Table 2.

Treatments	Protozoa (Log/ml) ^{ns}	Methanogen (x10 ⁵ cfu/ml)*	Decrease of methanogen (%)
PO	3.41 ± 0,03	2.90 ^d ± 3,00	-
P1	3.35 ± 0,11	2.30 ^c ± 4,24	20.68

Table 2. Total protozoa and methanogen in rumen fluid of sheep

P2	3.43 ± 0,04	2.37 ^c ± 1,26	18.28
Р3	3.35 ± 0,05	0.72ª ± 2,63	75.17
P4	3.31 ± 0,08	1.42 ^b ± 3,30	51.03
Р5	3.30 ± 0,05	$2.80^{d} \pm 12,73$	3.45

Note : ns = non significant, * = significantly affect (P<0,05), values bearing different superscript within column show significant difference (P<0,05)

The graph of total methanogen in the study is presented in Figure 3.

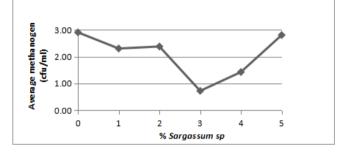


Figure 3. Mean of total methanogen in rumen fluid of sheep

The result of the present study showed that the supplementation of Sargassum sp. seaweed meal significantly affected (P<0.05) the decrease of total methanogen. On the contrary, Blanche et al [11] stated that the supplementation of 5% Ascophyllum nodosum brown seaweed did not affect the total methanogen in sacco. However, the result indicated that the supplementation of Ascophyllum nodosum could reduce the total methanogen by 89.90%, from 3.27 Log 103 x Δ CT with control feed to 0.33 Log 103 x Δ CT with 5% Ascophyllum nodosum. It indicated that phlorotannin in brown seaweed could affect the growth of methanogen. Shannon and Abdul-Ganam [12] stated that phlorotannin could bind bacteria protein like the enzyme produced by the bacteria and membrane cell that could produce lysis cells. Methanogen was expected to have the same reaction on phlorotannin as the other bacteria.

The result of study showed that the most significant decrease (75.17%) in total methanogen was obtained at 3% Sargassum sp. supplementation. Treatment with 0.03% monensin supplementation as the positive control could decrease 20.68% methanogen. It was assumed that 3% Sargassum sp. supplementation could lower more methanogen than the supplementation of 0.03% monensin. Morgavi et al. [13] stated that ionophore such as monensin was evidenced to inhibit the production of methane gas. Moreover, monensin can hinder Ruminococci but does not affect Fibrobacter succinogenes, so that monensin can be frequently used as anti-methanogen. Therefore, Sargassum sp. is the potential alternative to chemicals to decrease the amount of methanogen and methane gas emission.

The amount of methanogen in the treatment and the supplementation of 4% Sargassum sp. (P5) was relatively similar to the amount of methanogen in control feed (P0). It was expected that the higher the level of Sargassum sp., the lower phlorotannin efficacy to decrease methanogen, which indicated the change in ruminal microbe diversity. The higher the supplementation level, the lower the amount of fibrolytic which does not produce H2, and therefore the methanogen amount increased. It was in line with Morvan et al. [14], that the population of fibrolytic bacteria had a positive correlation with the amount of methanogen in the rumen of several cattle such as cow and sheep. It was supported by Wang et al [15], that supplementing 500 μ g/ml phlorotannin could affect the diversity of particular species of fibrolytic bacteria by decreasing the

amount of Fibrobacter succinogenes (H2-producing fibrolytic bacteria) but did not affect Ruminococcus flavefaciens (H2-producing fibrolytic bacteria). Morgavi et al [13] claim that increasing the number of bacteria that did not produce H2 like F. succinogenes could be the alternative to decrease the emission of methane gas without affecting fiber digestibility and to increase the efficiency of fiber digestibility my microbe. Patra et al [16] stated that the contributing factor to the ability of tannin to decrease the production of methane gas and the amount of methanogen was the difference in tannin structure where the condensed tannin had more significant potential. Jayanegara [17] stated that the structure of tannin compound is different across plant types which accounts for the non-toxic properties in several tannins at a certain level.

The result of the research indicated that the supplementation of 4% Sargassum sp. brown seaweed meal did not significantly (P>0.05) the population of protozoa. It was in contrast with the result of Belanche et al. [10] that the addition of 2% Ascophyllum nodosum brown seaweed in vitro could decrease protozoa activity up to 23%. Koivikko [18] informed that the abundance of phlorotannin in brown seaweed could reach 15% from the dry weight. The trend of total protozoa is presented in Figure 4.

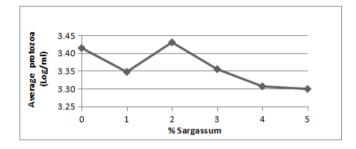


Figure 4. The mean of total protozoa in rumen fluid of sheep

Total dry matter of Sargassum sp. in this research was 8.12% and assumed to contain low phlorotannin and therefore less effective to decrease protozoa. Cahyaningrum et al [19] reported that the content of phlorotannin in seaweed Sargassum polycystum obtained from Poktunggal beach in Yogyakarta was 0.61±0.27 mg PGE/g extract. In this research, protozoa were presumed to adapt with the low phlorotannin.

The non-significant effect of the supplementation of Sargassum sp. on the population of protozoa may partly due to the 9-37% of protozoa that formed a symbiotic relationship with methanogen [4]. Phlorotannin does not explicitly lyse the membrane if protozoa like saponin. The mechanism of phlorotannin is similar to that of tannin in the plant in the land regarding binding and sedimenting the protein of bacteria cell which causes bacteria lysis [12]. Similarly, Laia [20] reported that protozoa population was not the dominant factor in methanogenesis because only a small fraction of methanogen which forms a symbiosis with protozoa; therefore when an inhibitor agent is given to the methanogen, it has a non-significant effect on the number of protozoa.

The supplementation of Sargassum sp. brown seaweed on the concentration of Volatile Fatty Acids (VFA) varied across treatments. The mean VFA (Table 2) of treatments R1, R2, R3, R4 and R5 was 190.5 mM; 200.5 mM; 227.5 mM; 240.5 mM and 172 mM, respectively. The data showed that treatment R4 (3% Sargassum sp.) had the highest VFA and R5 (4% Sargassum sp.) had the lowest VFA. Qomariyah et al [21] stated that the increase of total VFA was affected by the fermentability of feed by ruminal microbe to degrade the feed. The higher the degradation of carbohydrate and protein infeed, the higher the VFA and ammonia. The result of the analysis of variance showed that the treatments had a significant effect.

Prayitno et al [3] informed that the optimum level of VFA concentration for ruminal microbe growth was 80

- 180 mM. In the present study, treatments R1, R2, R3 and R4 was above the threshold. R1 consisted of 60% forage, and 40% concentrate feed without Sargassum sp. brown seaweed showed an excessive VFA. It indicated that treatment feed was easily fermentable in the rumen. Ramadhan et al [22] stated that the concentrate contained a highly digestible carbohydrate, so it was readily fermented by ruminal microbe into VFA.

The optimum level is presented from the curve graph (Figure 3). The polynomial orthogonal test indicated that 3% Sargassum sp. resulted in the highest VFA while the 2 % Sargassum sp. was the intersecting line between quadratic, cubic and quartic lines. The supplementation of 1-3% brown seaweed meal showed an increasing total VFA, partly due to the rising number of fiber-degrading bacteria because of the supplementation of Sargassum sp. brown seaweed. Wang et al [15] informed that the addition of phlorotannin to cattle beef would affect the total bacteria and cellulolytic bacteria.

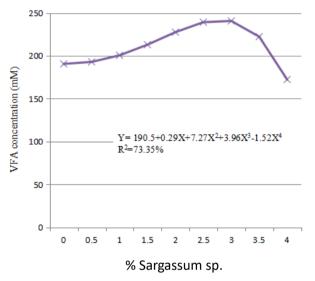


Figure 5. The curve of total VFA concentration (mM)

The highest production of VFA was at 2% supplementation of Sargassum sp. which decreased by 4% due to the decreasing activity of cellulolytic bacteria which lowered carbohydrate degradation. The supplementation of 2% Sargassum sp. was the optimum level because the range of normal VFA concentration in feed supplemented with 3% Sargassum sp. brown seaweed meal was not significantly different from that of 4%. Feed supplemented with 2% Sargassum sp. produced ±227.5 mM total VFA. It was following the hypothesis that 2% Sargassum sp. supplementation could increase total VFA. Belanche et al [10] informed that in vitro analysis using the rumen liquid of Friesian-Holstein cow supplemented with Ascophyllum nodosum brown seaweed showed a total VFA which tended to increase although not significant. The result of the supplementation of Ascophyllum nodosum 0%; 0.5%; 1%; 2% was 78.7 mM; 79.7 mM; 80.6 mM and 82.1 mM, respectively.

4. Conclusion

Supplementation of 2 % brown seaweed (Sargassum sp.) in sheep feed was an effective anti methanogenesis agent as evidenced by the production of total gas, methane gas, methanogen population, and protozoa population. Supplementation of 2% brown seaweed (Sargassum sp.) in sheep feed produces optimal volatil

fatty acids.

Acknowledgments

The authors thank the Higher Education, Directorate of Research and community Service, Ministry of Research, Technology and Higher Education, Republic of Indonesia for the research grant of Applied Research. Special thanks to Mrs Restu who provided valuable help in the Feedstuff Laboratory of Animal Science, Jenderal Soedirman University.

REFERENCES

- [1] Wuebbles D. J., Hayhoe K., "Atmospheric methane and global change," Earth-Science Rev, vol. 57, no. 3–4, 2002. DOI: 10.1016/S0012-8252(01)00062-9.
- [2] Sirohi S.K., Pandey N., Goel N., Singh B., Mohini M., Pandey P., Chaudhry P P., "Microbial Activity and Ruminal Methanogenesis as Affected by Plant Secondary Metabolites in Different Plant Extracts," Environ.
 Eng, 2009. https://www.researchgate.net/publication/215876927_Microbial_Activity_and_Ruminal_methanogenesis_ as_affected_by_Plant_Secondary_Metabolites_in_Different_Plant_Extracts (accessed Jan. 18, 2021).

[3] Prayitno C. H., Subagyo Y., Suwarno, "Supplementation of sapindus rarak and garlic extract in feed containing adequate Cr, Se, and Zn on rumen fermentation," Media Peternak, vol. 36, no. 1, 2013. DOI: 10.5398/medpet.2013.36.1.52.

- [4] Newbold C. J., Lassalas B., Jouany J. P., "The importance of methanogens associated with ciliate protozoa in ruminal methane production in vitro," Lett. Appl. Microbiol, vol. 21, no. 4, 1995. DOI: 10.1111/j.1472-765X.1995.tb01048.x.
- [5] Inthapanya S., Preston T. R., Leng R. A., "Mitigating methane production from ruminants; effect of calcium nitrate as modifier of the fermentation in an in vitro incubation using cassava root as the energy source and leaves of cassava or Mimosa pigra as source of protein," Livest. Res. Rural Dev, vol. 23, no. 2, 2011. https://www.researchgate.net/publication/287593155_Mitigating_methane_production_from_ruminants_ effect_of_calcium_nitrate_as_modifier_of_the_fermentation_in_an_in_vitro_incubation_using_cassava_ro ot_as_the_energy_source_and_leaves_of_cassava_or_Mimosa (accessed Jan. 18, 2021).
- [6] Ogimoto K., Imai S., "Atlas of Rumen Microbiology, " Tokyo: Japan Scientific Societies Press, 1981. https://www.worldcat.org/title/atlas-of-rumen-microbiology/oclc/7921878 (accessed Jan. 18, 2021).
- [7] Paynter M. J. B., Hungate R. E., "Characterization of Microbacterium mobilisi Isolated from Bovine Rumen," J.
 Bact, no. 95, pp. 1943–1951, 1968. https://jb.asm.org/content/95/5/1943.short (accessed Jan. 18, 2021).
- [8] Calinski T., Steel R. G. D., Torrie J. H., "Principles and Procedures of Statistics: A Biometrical Approach.," Biometrics, vol. 37, no. 4, 1981. DOI: 10.2307/2530180.
- [9] Ellis, J. L., Dijkstra J., Kebreab E., Bannink A., Odongo N. E., McBride B. W., France J., "Aspects of rumen

microbiology central to mechanistic modelling of methane production in cattle," Journal of Agricultural Science, vol. 146, no. 2. 2008. DOI: 10.1017/S0021859608007752.

- [10] Belanche A., Ramos-Morales E., Newbold C. J., "In vitro screening of natural feed additives from crustaceans, diatoms, seaweeds and plant extracts to manipulate rumen fermentation," J. Sci. Food Agric, vol. 96, no. 9, 2016. DOI: 10.1002/jsfa.7481.
- [11] Belanche A., Jones E., Parveen I., Newbold C. J., "A metagenomics approach to evaluate the impact of dietary supplementation with Ascophyllum nodosum or Laminaria digitata on rumen function in Rusitec fermenters," Front. Microbiol, vol. 7, no. MAR, 2016. DOI: 10.3389/fmicb.2016.00299.
- [12] Shannon E., Abu-Ghannam N., "Antibacterial derivates of marine algae: An Overview of Pharmacological Mechanisms and Applications," Mar. Drugs, vol. 14, no. 81, pp. 1–23, 2016. DOI: doi:10.3390/md14040081.
- [13] Morgavi D. P., Forano E., Martin C., Newbold C. J., "Microbial ecosystem and methanogenesis in ruminants," Animal, vol. 4, no. 7, 2010. DOI: 10.1017/S1751731110000546.
- [14] Morvan B., Bonnemoy F., Fonty G., Gouet P., "Quantitative determination of H2-utilizing acetogenic and sulfate-reducing bacteria and methanogenic archaea from digestive tract of different mammals," Curr. Microbiol, vol. 32, no. 3, 1996. DOI: 10.1007/s002849900023.
- [15] Wang Y., Alexander T. W., Mcallister T. A., "In vitro effects of phlorotannins from Ascophyllum nodosum (brown seaweed) on rumen bacterial populations and fermentation," J. Sci. Food Agric., vol. 89, no. 13, 2009. DOI: 10.1002/jsfa.3717.
- [16] Patra A., Park T., Kim M., Yu Z., "Rumen methanogens and mitigation of methane emission by anti-methanogenic compounds and substances," Journal of Animal Science and Biotechnology, vol. 8, no. 1. 2017. DOI: 10.1186/s40104-017-0145-9.
- [17] Jayanegara A., "Reducing methane emissions from livestock: nutritional approaches," Proceedings of Indonesian Students Scientific Meeting (ISSM), Institute for Science and Technology Studies ISTECS) European, pp. 18-21, 2008.
- [18] Koivikko R., "Brown Algae Phlorotannins Improving and Applying Chemical Methodsi," Turku, Finland., 2008. http://www.doria.fi/handle/10024/36054 (accessed Jan. 18, 2021).
- [19] Cahyaningrum K., Husni A., Budhiyanti S. A., "Activity of Brown seaweed (Sargassum polycystum) extract (Abstract in English).," Agritech, vol. 36, no. 2, pp. 137–144, 2016. DOI: https://doi.org/10.22146/agritech.12857.
- [20] Laia N., "Effectivity of herbal plant on rumen fermentation, methane gas and protozoa population by in vitro," Bogor Agricultural University, 2016.

- [21] Qomariyah N., Retnani Y., Jayanegara A., Wina E., Permana I. G., "Influence of Biochar and Liquid Smoke Additives from Cacao-Pod Husks on In vitro Ruminal Fermentation Characteristics," Adv. Anim. Vet. Sci., vol. 9, no. 4, 2021. DOI: 10.17582/journal.aavs/2021/9.4.533.543.
- [22] Ramadan M. F., Hassan N. A., Elsanhoty R. M., Sitohy M. Z., "Goldenberry (Physalis peruviana) juice rich in health-beneficial compounds suppresses high-cholesterol diet-induced hypercholesterolemia in Rats," J. Food Biochem., vol. 37, no. 6, 2013. DOI: 10.1111/j.1745-4514.2012.00669.x.