

Anticancer Capability of Go-Ark With *Plectranthus Amboinicus* Against Hepatocellular Carcinoma

Vinotha M¹, Kamalaveni J¹, Nithya V*

*¹Department of Animal Health and Management, Pharmacognosy Lab, Alagappa University, Karaikudi-630 003, Tamil Nadu, India.

Corresponding author: Nithya V

E-mail: nithyav@alagappauniversity.ac.in

Abstract

Cancer is a life threatening disease and an emerging public health issue that affects humans worldwide. The main objective of the present investigation is to detect the effects of Cow ark, synergistic with ethanolic leaves extract of *Plectranthus amboinicus* on Diethylnitrosamine (DEN) and 2-Acetylaminofluorene (AAF), stimulated hepato carcinogenesis. Cow ark and 5% Ethanolic extract of *P. amboinicus* were systematically prepared. Identification of *in situ* DNA fragmentation was performed by TUNEL assay to explore apoptogenic characteristics of distillate with *P.amboinicus* leaves extract. Apoptotic study observed that a maximum number of apoptotic cells were detected in the liver cells belong to (AB) group, when compared with other tested groups. The Go-ark with *P.amboinicus* extract also showed chemo preventive activities by curing hepatocarcinogenesis activated by DEN/AAF carcinogens. This study proved to be a promising anticancer effect of distillate and *P. amboinicus* extract synergistically in increasing programmed cell deaths.

Key Words: Plectranthus amboinicus, apoptosis cells, hepatocarcinoma cells, in vivo

INTRODUCTION

Cancer is still a challenging disease and a major public health problem that affects individuals worldwide. Cancer stands as the second major cause of mortality in the United States. Recent data alarming that projected new cancer occurrences and mortality in the United States in 2022, offers a complete view of development of cancer. New cases are determined to increase by 70% in the forthcoming two decades. A statistical report reveals that in 2022, 609,360 people in the US are expected to die of cancer, accounting for 1700 deaths per days [1-4].

Hepatocellular carcinoma (HCC) is one of the most common malignancies reported in the world. Due to the rapid global pandemic of hepatitis B and C viral infections, the occurrence of HCC is known to be increasing in Asia and the majority of Western countries [5-8]. The treatment for advanced HCC is found to be very limited and novel approaches are essential [9-10].

Several investigations described that the traditional organic and animal origin products can be used for improved general health [11-12]. A study conducted by Cow Science Research Center reported that indigenous cow breed urine contains, therapeutic values in treating various kinds of HCC malignancy, Protective effects of Cow urine distillate such as, declining the enzymatic actions of SGOT, SGPT, ALP, GGT and total bilirubin with distillate tends to reduce the enhanced enzyme levels, indicating that it struck the action of free radical generations [13-17].

P. amboinicus is also a prolific plant used in Asia and African countries. This species was well recorded, for their use in curing, many ailments for human health A recent study elucidated that methanolic leaf extracts of *P. amboinicus* exerted a vital role in the management of oxidative stress, using their antioxidant mechanisms [17-18]. Another study in the existing line, revealed that hydro alcoholic extract of *P. amboinicus* was reportedly exhibited anti- inflammatory and anti-tumor activities [19]. Therefore, in the

present investigation was performed aiming at, to find the anti-cancer efficacy of Cow urine distillate, combined with *P. amboinicus* ethanolic extract in the hepatocytes of HCC cells *in vivo*.

MATERIALS AND METHODS Collection of Cow urine

The fresh and very first flow of urine was collected in a container, a wide conical flask, from a healthy breed, Bos indicus, in the early morning. Collected urine was brought to the nearby laboratory, as early as, and filtered carefully, using Whatmann filter paper.

Preparation of Go-ark

As collected sample was subjected to the simple distillation set up for the urine distillation process. A wide mouth conical flask with broad rounded bottom, was placed on a heating mantle, and allow continuously, then it was linked with water cooled condenser, at one end, while, other side was connected with a beaker, which was incorporated into a large container for the distillate collection [19]. Clean filtrate distillate was received in a sterile container carefully, and it was kept as a stock solution. From this stock distillate, a varied concentration, such as 5% was formulated to detect anticancer activity potentials.

Collection of liver tissues

The rats were stimulated with carcinoma, by injecting 200 mg/kg-1 Diethyl Nitrosamine (DEN) dissolved in corn oil as vehicle, followed by a recovery of 15 days as food, blended with 2- acetylaminofluorene (0.02% AAF) as enhancer of hepatocarcinogenesis. At the end of the treatment tenure, the rats were weighed, and subsequently sacrificed by using ether anesthesia. Liver were detached as quick, and washed well in 1.15 concentrated potassium chloride solution suddenly. As processed liver samples were kept in care for Tunel assay and slices were viewed under confocal microscope for critical analysis.

TUNEL Assay (Apoptotic observations)

TUNEL is a dynamic staining method for detecting dead cells. It has been demonstrated as a powerful tool for determining manifestations of cell death in biological systems, in particular, while used as part of a panel of complementary biomarkers. Terminal deoxynucleotidyl transferase – aided dUTP nick end labeling the free S -hydroxy terminal. Tissues isolated from the rats of each group were cut into pieces from the largest lobe and were well fixed in 4% paraformaldehyde, developed in paraffin blocks, subsequently, sections were done at dead end. A TUNEL fluorometric system was performed, based on the manufacturer instructions which was displayed. Such stained slides with TUNNELS were brought under fluorescent microscope. Prepared slides were tested and a Scoring measurement was also carried out, at x200 magnifications and chosen 5 location spots, randomly [20]. In each spot, Green coloured cells were enumerated as TUNEL positive apoptotic cells of examined tissues. Fluorescence images obtained, using fluorescence imaging systems.

Statistical application

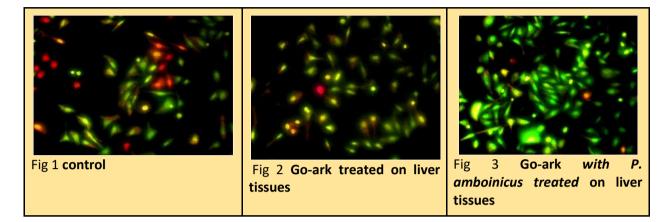
The values expressed in the graph as mean + or -SD. By applying SPSS software package, ANOVA test was performed to know if the statistically significant difference between varied groups at p<0.05 were determined.

RESULTS

Detection of Apoptosis cells, using TUNEL system

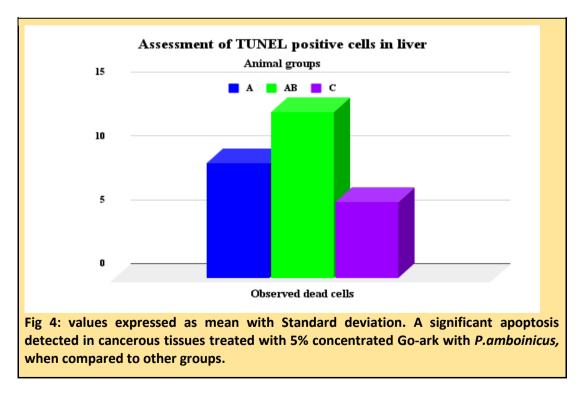
The tested and stained liver tissues of DEN/AAF stimulated rats, exhibited that a small number of fluorescent Green stained cells were detected, whereas, Red and Orange colored stained cells were

declared as viable cells, still present time. Our TUNEL staining results showed the presence of apoptotic cells with abundant green fluorescence of liver slices, which were received from DEN/AAF activated rats, treated with 5% Cow urine distillate (Fig-1-3). The present observation is obviously evident that many prominent intensified signals of DNA breaks were clearly appeared, while, distillate along with extracts of *P. amboinicus*, treated liver cells, in specific, HepG2 cells, when compared to vehicle control group (C) rat liver cells.



MEASUREMENT OF TUNEL POSITIVE CELL SCORING

Counting of dead cells (apoptotic cells) was also performed by using TUNEL positive cells score. Results obtained from TUNEL assay, subsequently, positive cells scoring were determined. The findings were depicted in the graphical representation as in Fig 4 which reveals that there was a significant difference between Distillate with *P. amboinicus* extract treated group with other designed rat groups at p<0.05% in apoptotic cells detected.



DISCUSSION

The rational formulation of therapeutic entities from traditional medicine to treat dreadful diseases and pathogenic infections since Rig Veda and it was compared as nectar [21-23]. Cow urine acts as a 'bio

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enhancer' to the specific antibiotics previously reported. The different fractions of cow urine distillate have been reported to be antimicrobial, anticancer and larvicidal properties. Existing lines of studies shown that cow urine distillate is an immune enhancer [24- 31]. Moreover, cow urine possess ,antioxidant characteristics and helps to lymphocytes to survive by suppressing, their suicidal cells and repairing the injured DNA. Hence, cow urine distillate used as effective anti cancer therapy [32-36]. Cow urine was observed in a study, as a chemopreventive ability and resulted, the incidence of tumor was statistically less was recorded [37-41]. However, cow urine distillate affects the other disorders or other cancer related have never been highlighted in earlier investigations. Therefore current study conducted, as a complementary investigation, for its anti cancer effects of cow urine distillate along with ethanol extract of *P. amboinicus in vivo*, in using hepatocarcinogenesis models. This would offer effective and authentic information to Cancer research.

Apoptosis is associated either with external stimulus or stress inducing agent of pathological consequences. In chemotherapeutic treatments, it was reported to induce apoptotic mechanisms of tumor cell death, and it could be a positive means for present cancer treatment [42-43].) Besides, apoptosis is an innovative target for removing cells which are transformed into advanced stages, whereas other practiced mechanisms exhibited were unsuccessful to block the upstream cancer development [44-45], The therapeutic activation of *P. amboinicus* has been analyzed by many research groups and reported to have strong cytotoxic and anti tumor potentials [46].

In several investigations, identification of chemo preventive stimulated a programmed cell death in HCC in animal models were demonstrated, including Humans [47-49]. Analyzing DNA fragments *in situ*, using TUNEL assay is a popular technique to detect apoptosis and its visualizations, labeling of DNA fragments, due to apoptosis [50]. In earlier stages of liver carcinoma, apoptosis, extremely compensate, cell proliferation [51].

Quantitative assessment of apoptosis used to reveal the tendency to enhance in the conversion of normal rat apoptosis to malignant cells. In our investigation variation in difference in TUNEL apoptotic positive cells were found to be markedly different (p < 0.05) between control and cancer stimulated rat groups.

Moreover, a good agreement with previous studies, such as Hae-Jeung et al, [52] described that supplementation of *P. amboinicus* with distillate have shown to be a significant anti cancer effects against DEN/AAF stimulated HCC, were ideal by TUNEL positive cells count in the examined liver slices. Our findings showed that an abundant number of death cells were observed in the treatment of go-ark with ethanol extract of *P. amboinicus* than other treatment schedules. This is apparent evidence that pre-neoplastic cells, activated by DEN/AFF are more vulnerable to the apoptotic cells, when compared to normal cells. This might be attributed to the synergistic effects of cow urine distillate and *P. amboinicus* ethanol extract, in killing cancer cells in the treatment. Interestingly, normal cells were not affected by the distillate and extract treatment. These findings proved that distillate and *P. amboinicus* extract altogether, showed notable hepatoprotective effects in the liver cells [53]..

CONCLUSION

Above comprehensive study affirmed that Go-ark extract of *P. amboinicus* played a significant role in increasing apoptosis of cancer cells in stimulated liver carcinogenesis.

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References

- 1. Rebecca L. Siegel MPH,Kimberly D. Miller MPH,Hannah E. Fuchs BS,Ahmedin Jemal DVM, PhD;Cancer statistics, 2022: A cancer journal for clinicans ; Volume 72 issue 1, p no7-31
- 2. Seeff , L.B. and J.H. Hoofnagle , 2006. Epidemiology of hepatocellular carcinoma in areas of low hepatitis B and hepatitis C endemicity . Oncogene , 25 (27) : 3771-377.
- 3. Bruix , J. , A.J. Hessheimer , A. Forner , L. Boix , R. Vilana and J.M. Llovet , 2006. New aspects of diagnosis and therapy of hepatocellular carcinoma . Oncogene , 25 (27) : 3848-3856 .
- 4. Nithya V. (2021) Indigenous cow ark with Allium Sativum -A key to therapeutic and an effective antioxidant to hepatocellular carcinoma, International Journal of Aquatic Science 12 (2), 1631-1644.
- 5. Thavasuraj S, Nithya V. (2020). Evaluation on efficacy of fatty acids and conjugated Linoleic Acid [CLA] derived from indigenous cow milk Against hepato cellular carcinoma : A key and novel approach, Biochemical and cellular archives, 20(2), 3253-3257.
- 6. Thavasuraj S, Nithya V, Vinotha M, Chinniah S, Archunan G. (2020). Assessment of dosage response of cow milk fat isomers against hepatocellular carcinoma cell lines : A key to novel approach for anticancer. Biochemical and cellular archives, 20(2), 3223-3227.
- 7. Silambarasan V, Deepalakshmi G, Sankarganesh D, Nithya V, Archunan G.(2020). Identification of potential pheromone source in sows. Behavioural processes, 168, 103940.
- 8. Bhaskar, A., Nithya, V., & Vidhya, V.G. (2011). Phytochemical evaluation by GC-MS and antihyperglycemic activity of Mucuna pruriens on streptozotocin induced diabetes in rats. Journal of Chemical and Pharmaceutical Research, 3(5), 689-696.
- 9. Nithya, V. (2021). In vitro antibacterial, antioxidant and time-kill kinetics of cow ark with Plectranthus Amboinicus extract against human pathogens. International Journal of Aquatic Science, 12 (1), 314-321.
- 10.Nithya, V. (2021). Indigenous Cow Ark with Allium Sativum -A Key To Therapeutic And An Effective Antioxidant To Hepatocellular Carcinoma. International Journal of Aquatic Science, 12 (2), 1631-1644.
- 11.Gururaja MP, Joshi AB, Joshi H, Sathyanarayan D, Subrahmanyam EVS, Chandrashekhar KS. Attenuation of Carbon Tetrachloride -induced hepatotoxicity by cow urine distillate in Rats. Biomed Environ Sci. 2009; 22:345-347.
- 12. Chauhan RS, Garg N. Cow Therapy as an Alternative to Antibiotic. Banglore, Karnataka: Indian Science Congress; 2003.
- 13.Ramachandran R, Vinothkumar A, Sankarganesh D, Suriyakalaa U, Aathmanathan VS, Kamalakkannan S, Nithya V, Angayarkanni J, Archunan G, Akbarsha MA, Achiraman S. (2020). Detection of estrous biomarkers in the body exudates of Kangayam cattle (Bos indicus) from interplay of hormones and behavioral expressions, Domestic animal endocrinology, 72, 106392.
- 14.Balu Krishnakumar, Ravikumar, S., Pandiyan, V., Nithya, V., Sylvestre, S., Sivakumar, P., Surya, C., Agnel, Arul John, N., & Abilio JFN Sobral. (2020). Synthesis, characterization of porphyrin and CdS modified spherical shaped SiO2 for Reactive Red 120 degradation under direct sunlight. Journal of Molecular Structure, 1210, 128021.
- 15.Hongru Zhang, Joe Antony Jacob, Ziyu Jiang, Senlei Xu, Ke Sun, Zehao Zhong, Nithya, V., & Achiraman Shanmugam. (2019). Hepatoprotective effect of silver nanoparticles synthesized using aqueous leaf extract of Rhizophora apiculata. International journal of nanomedicine, 14, 3517.
- 16.Ravikumar, S., Pandiyan, V., Manawwer Alam, Naushad Ahmad, Nithya, V., Balu Krishnakumar, & Abilio.J.F.N. Sobral. (2021). Costus speciosus koen leaf extract assisted cs-znx (X= O or S) nanomaterials: Synthesis, characterization and photocatalytic degradation of rr 120 dye under uv and direct sunlight. Journal of Molecular Structure 1225, 129176.
- 17.Vinotha, M., &Nithya, V. (2021). Evaluation of cow ark enhanced Plectranthus Amboinicus for the potential of antioxidant, antimicrobial, and larvicide potentials –in vitro. International Journal of Aquatic Science, 12(2), 1840-1848.
- 18.Vinotha, M., &Nithya, V. (2021). Potential hepatoprotective of cow ark with Plectranthus Amboinicus against changes in the level of CCL4-induced liver intoxication and antioxidant enzymes. Annals of the Romanian Society for Cell Biology, 25 (4) 19415-19422.

- 19. Vinotha M, Thavasuraj S, Chinniah S, Nithya V. (2020). Antimicrobial, Antibiofilm and Antioxidant Effects of Medicinal Plants Extract with Indigenous Cow Ark Against Human Pathogens. International journal of advanced science and technology, 29(3),569-583.
- 20.Silambarasan V, Gayathiri S, Deepalakshmi D, Shahitha Banu, Nithya V, Archunan G.(2020) Cloning and sequencing of α -2u globulin of rat preputial gland to assess its longevity in the context of developing an effective rodent trap. Indian Journal of Biochemistry and Biophysics, 56(6),433-438.
- 21.Nithya V. (2015). Evaluation of antidiarrheal activity on Coriandrum sativum linn., in wistar albino rats. World Journal of Pharmaceutical research, 4(5),638-643.
- 22.Nithya V.(2011). Anti-inflammatory activity of Lawsonia ulba Linn., in wistar albino rats. Asian J Sci Tech. 2011;4:001–3.
- 23.Nithya V, Baskar A. (2011). A preclinical study on wound healing activity of Lawsonia ulba Linn, Research Journal of Phytochemistry,5(2)123–129, 2011.
- 24.Randhawa GK, Kullar JS, Rajkumar. Bioenhancers from mother nature and their applicability in modern medicine. Int J Appl Basic Med Res 2011;1:5-10
- 25.Dutta D, Devi SS, Krishnamurthi K, Chakrabarti T. Anticlastogenic effect of redistilled cow's urine distillate in human peripheral lymphocytes challenged with manganese dioxide and hexavalent chromium. Biomed Environ Sci 2006;19:487-94.
- 26.Nithya, V. (2011). Evaluation of the wound healing activity of datura metel linn., in wistar albino rats, Inventi Rapid: Planta Activa, 2(1).
- 27.Nithya, V., Brinda, P., & Anand, K.V. (2011). Wound healing activity of leonotis nepetaefolia R.Br., in wistar albino rats. Asian Journal of Pharmaceutical and Clinical Research, 4(2), 23-26.23.
- 28.Nithya, V., & Balasubramanian, K. (2008). Evaluation of wound healing activity of Polygonum barbatum Linn in wistar albino rats. Research journal of Biological sciences, 1 (1), 14-20.
- 29.Nithya, V. (2011). Phytochemical studies of lawsonia ulba linn.,-a medicinal plant. Inventi Rapid: Ethnopharmacology, 2(1).
- 30.Nithya, V., & Anusha, B. (2011). Evaluation of phytochemicals studies on polygonum barbatum linn. Inventi Rapid: Ethnopharmacology, 2(1)
- 31.Nithya, V. (2011). A review of pharmacological studies of some medicinal plants as antimicrobial and feed additives. Inventi Rapid: Ethnopharmacology, 2(1).
- 32.Nithya V, Baskar A (2011). A preclinical study on wound healing activity of Lawsonia ulba Linn, Research Journal of Phytochemistry,5(2)123–129, 2011.
- 33.Nithya V.(2011). Anti-inflammatory activity of Lawsonia ulba Linn., in wistar albino rats. Asian J Sci Tech. 2011;4:001–3.
- 34.Nithya, V. (2011). A review on clinical uses of aloe vera. Inventi Rapid: Pharmacy Practice, 2(2).
- 35.Nithya, V. (2011). A review on treating sickle cell disease through indian medicinal plants. Inventi Rapid: Ethnopharmacology, 2(2).
- 36.Nithya, V. (2011). Anti-inflammatory activity of allium sativum linn., in wistar albino rats. Inventi Rapid: Ethnopharmacology, 2(1).
- 37.Nithya, V. (2014). Phytochemical and wound healing activity of Allium Sativam linn., in wistar albino rats. World Journal of Pharmaceutical Research, 3 (7), 491-498.
- 38.Nithya, V. (2015). Evaluation of antidiarrheal activity on Coriandrum sativum Linn. in wistar albino rats. World Journal of Pharmaceutical Research, 4, 638-43.
- 39.Nithya, V. (2015). Evaluation of phytochemicals studies on Coriandrum sativum Linn. World Journal of Pharmaceutical Research, 4 (5), 1465-1471.
- 40.Nithya, V., Anusha, B., & Muhammadllyas, M.H. (2014). Antimicrobial activity of certain antibiotics on the microbial isolates of keratitis patient. World Journal of Pharmaceutical Research, 3(10), 1226-1234.
- 41.Nithya. V. (2018). Bioactive Compound Analysis of Coriandrum Sativum L against Microbial Keratitis. Ophthalmology. Breakthroughs in Research and Practice, 268-282.
- 42.Nithya, V., & Bhaskar, A. (2014). A Review of Microbial Keratitis. World Journal of Pharmaceutical Research, 3 (10), 189-201.
- 43.Thavasuraj, S., &Nithya, V. (2021). In vitro anticancer potential of Anisomeles malabarica against cervical cancer cells. Annals of the Romanian Society for Cell Biology, 25 (4) 19423-19428.

- 44.Nithya, V. (2019). SubmitoLoc Identification of mitochondrial sub cellular locations of proteins using support vector machine, Bioinformation 15 (12), 863-868.
- 45.Nithya, V. (2014). Phylogenetic analysis of 16srdna and 18srdna sequences of bacteria and fungi from keratitis patients. World Journal of Pharmaceutical Research, 3 (10), 932-941.
- 46.Bhaskar, A., &Nithya, V. (2013). In silico Structural Analysis of 16S rDNA Sequences of Bacteria Isolated from Keratitis Patients. Trends in Bioinformatics, 6 (2), 45-61
- 47.Pugalenthi, G., Nithya, V., Chou, K.C., & Archunan, G. (2020). Nglyc: A random forest method for prediction of N-Glycosylation sites in eukaryotic protein sequence, Protein and Peptide Letters, 27 (3), 178-186.
- 48.Kerr JF, Winterford CM & Harmon BV 1994 Apoptosis. Its significance in cancer and cancer therapy. Cancer 73 2013–2026.
- 49.Norazsida, R., Pakeer, O. &Taher, M., 2017. The antimalarial properties of essential oils of the leaves of Malaysian Plectranthus amboinicus (Lour) spreng in mice infected with Plasmodium berghei. International Medical Journal Malaysia, 16 (1), pp.67–74.
- 50.Kerr, J.F., C.M. Winterford and B.V. Harmon, 1994. Apoptosis: It is significant in cancer and cancer therapy. Cancer, 73: 2013-2026.
- 51.Schulte-Hermann, R., W. Bursch, B. Grasl-Kraupp, L. Torok, A. Ellinger and L. Mullauer, 1995. Role of active cell death (apoptosis) in multistage carcinogenesis. Toxicol. Lett., 82-83: 143-148.
- 52.Hae-Jeung, L., L. Sang-Ah and C. Haymie, 2005. Dietary administration of inositol and/or inositol-6-phosphate prevents chemically-induced rats' hepatocarci-nogenesis. Asian Pac. J. Cancer Prev., 6: 41-47.
- 53.Suresh Kumar, S.V. and S.H. Mishra, 2008. Hepatoprotective effect of Pergularia daemia (Forsk.) ethanol extract and its fraction. Indian J. Exp. Biol., 46: 447-452.